

## BRIEF PROFILE OF DOCTOR TREE (*MORINGA OLEIFERA* LAM.)

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### ABSTRACT

The paper presents a brief profile of Doctor Tree (*Moringa oleifera* Lam.) - including phytochemical constituents, minerals, vitamins, aminoacids, fatty acids, nutritional and pharmaceutical properties of its various morphological components, seed-oil and exudates (gum). Its antioxidant, antidiabetic, anti-atherosclerotic, antimicrobial and nematocidal activities and its effectiveness in many diseases, frequently reported in literature, have been described. Its role in mitigating climate change and global warming and efficacy against *Anopheles gambiens* and *Aedes aegyptii* is also reported in literature. *M. oleifera* contains some 163 phytochemicals of which 40 are phenolics and flavonoids and it is active in more than 80 diseases including neuro-pharmacological effects against Alzheimer's disease, anticancer and antitumour activity, wound closure and tissue regeneration, anti-urolithiasis, testicular damage-repairing, antihypertensive, anti-ulcerogenic, radio-protector etc. It is a nice fodder and suggested for poverty alleviation in poor countries.

**Key-words:** *Moringa olifera* Lam., phytochemicals, nutritional, environmental and medicinal importance.

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### INTRODUCTION

Doctor tree (*Moringa oleifera* Lam.), belongs to Family Moringaceae. The genus name comes from "Murungai" (Tamil) means twisted pod and species name is from Latin words "Oleum" (oil) and "ferre" (to bear). It is a common cultivated rapid-growing tree. There are some 126 vernacular names of the plant (Shamim *et al.*, 2018) – Sohanjna (saijan), Swenjera (Sindhi), Doctor Tree, Arengé Behan, Clarifier tree, Drumstick tree, Horseradish tree, Benzoline tree, Ben oil tree, Super food tree, Miracle tree, Tree of life, Moringa, etc. Native to Indo-Pak sub-continent. Widely distributed in many countries for its nutritional and medicinal importance – India, Pakistan, Bangladesh, Afghanistan, South-East Asia, Philippines, Indonesia, Taiwan, Caribbean, Africa, and America. The history of *M. oleifera* dates back to 150 BC. Dhakar *et al.* (2011) have reported that ancient Mauryan warriors of India were fed with Moringa leaf extract for extra energy.

*M. oleifera* grows from Sea level to 1400m. It coppices well (Shamim *et al.*, 2018). It can survive in very dry arid deserts with less than 400 mm annual rainfall (iwacu- Burundi, org/englishnews/Moringa-the-never-die-tree) and under various stressful conditions (Mahmood *et al.*, 2010). It has quite wide ecological amplitude. Sometimes referred to as "never die" tree (Swati *et al.*, 2018). It requires loamy and sandy soil (Thurber and Fahey, 2010). As regard to its growth, pollarding, coppicing and lopping or pruning are recommended for encouraging branching. (Denton and Grubben, 2004). Khan *et al.* (2021) have described its foliar micromorphology of its seedlings. It is a very useful plant, and every part of the plant is used by the mankind for millennia (Martin *et al.*, 2013), one way or the other (Abd Rani *et al.*, 2018) and in more than 80 countries (Mahmood *et al.*, 2010). The wood may be used for fire and poor-quality charcoal and in making paper pulp. A blue dye is produced from wood in Senegal and Jamaica (Halder and Kosankar, 2017). The oil extracted from nuts (seeds) is called 'Ben' or Behen' oil used as lubricant, in cosmetics and applied in rheumatism. The seeds are considered antipyretic. They are oleaginous and contain 38.4 % protein and 34.7 % fatty oil. The seed cake is used as fertilizer (NISC, 1998). *M. oleifera* is a miracle plant (Padayachee and Baijnath, 2012). It is good food and one of the best food supplements. It contains all essential amino acids (unusual for a plant) (<http://muneilla.wordpress.com/2012/04/16/moringa-global-warming-and-you/>). It is a fertilizer, efficient fuel, livestock feed and extremely safe edible plant (Mahmood *et al.*, 2010). There are many uses of *M. oleifera* in human and animal nutrition (Hanuk, 2018). Ganatra-Tejas *et al.* (2012) have reviewed from various publications that *M. oleifera* have very many traditional and pharmacological characteristics. It improves soil and provides raw materials for food, purification of water and cosmetics (Velázquez-Zavala *et al.*, 2016). Its wood yields blue dye (Misra *et al.*, 2011). A brief profile of *M. oleifera* benefits is presented in this paper.

## MATERIALS AND METHODS

Hundreds of research papers have so far been published on nutritional, biological, traditional, phytochemical, pharmaceutical, pharmacological and other characteristics of *M. oleifera* (to cite a few: Das *et al.*, 1957; Delaveau and Baiteau, 1980; Villasenor *et al.*, 1989a and b; Faizi and coworkers, 1994; 1997; Muyibim and Evison, 1994; Gillani *et al.*, 1994; Gassenschmidt *et al.*, 1995; Pollard *et al.*, 1995; Makkar and Becker, 1996; 1997; 1999; Njoku and Adikwu, 1997; Ramachandran *et al.*, 1980; Foidl *et al.*, 2001; Rao *et al.*, 2001; Anonymous, 2003; Ganatra–Tejas *et al.*, 2012; Tahir *et al.*, 2017; Mehta *et al.*, 2003; Annongu *et al.*, 2014; Luqman *et al.*, 2012; Alegbeleye, 2018; Agbogidi and Ilonda, 2012; Anwar and Rashid, 2007; Anwar *et al.*, 2007; Kalogo *et al.*, 2000; Kasolo *et al.*, 2010; Moyo *et al.*, 2011, 2012; Dhakar *et al.*, 2011; Nda Bigengeser and Narasiah, 1998; Fahey, 2005; Manzoor *et al.*, 2007; Isitua *et al.*, 2015; Stevens *et al.*, 2013; Martin and coworkers, 2013; Charoensin, 2014; Velázquez-Zavala *et al.*, 2016); Padayachee and Baijnath, 2012; Iqbal and Bhangar, 2006; Atawodi *et al.*, 2010; Sidhuraju and Becker, 2003; Bennet *et al.*, 2003; Torres-Castillo *et al.*, 2013; Razis *et al.*, 2014; Uniugbe *et al.*, 2014; Leone *et al.*, 2015; Guevara *et al.*, 1996, 1999; Mehta *et al.*, 2011a; Mishra *et al.*, 2011; Rani and Arumugam, 2017; Hussain *et al.*, 2014; Swathi, 2016; Igado and Oladale, 2016; Harimalala *et al.*, 2017; Maqsood *et al.*, 2017; Kumar, 2017; Singh, 2017; Shamim *et al.*, 2018; Abd Rani and associates, 2018; Singh *et al.*, 2019; Mahmood *et al.*, 2010; Gopalakrishnan *et al.*, 2016; Padmalochana, 2018; Shanmugavel *et al.*, 2018; Mahfuz and Piao, 2019; Pandey *et al.*, 2019; Aleksic, 2020; González – Romero *et al.*, 2020; Subramonie *et al.*, 2020, Srivastava *et al.*, 2020; Bekoe, 2020; Almeida *et al.*, 2021; Bhalla *et al.*, 2021; Lui *et al.*, 2021; Fidrianny and coworkers, 2021; Prithiviraj and Sumathy, 2021; Rubio-Sanz *et al.*, 2021; Basillo-Heradia and Gutierrez-Grijalava, 2022 (a very informative book on biological and pharmacological aspects of genus *Moringa* such as antioxidant, anti-inflammatory, antidiabetic properties, phytochemical and bioactive characteristics, genetic diversity and agronomy) and many more).

This paper on profile of doctor tree was prepared on consultation of some 300+ publications on the subject characteristics.

## BRIEF PROFILE OF *MORINGA OLEIFERA* LAM.

### 1) Phytochemicals

Many workers have studied phytoconstituents of *M. oleifera*. Garga *et al.* (2019) have reviewed and described the presence of flavonoids, tannins, saponins, cardiac glycosides, steroids, alkaloids, volatile oils, Anthraquinones, saponins glycosides, in leaves and seeds extract of this plant. Bhalla *et al.* (2021) identified 25 phytochemicals in aq. extract and 54 phytochemicals in methanolic extract of *M. oleifera* leaves. Regarding the presence of Anthraquinones the reports are controversial. Anthraquinones were reported to be completely absent from the leaf extracts by Kwaghe and Ambali (2009) in fresh and dry leaf samples. When extracted with chloroform, ethyl acetate or n-Butanol and Aqueous and Aqueous + Methanol extracts of leaves collected from Kibwezi, Makeuri County of Kenya (Okumu *et al.*, 2016) and Gujrat, India (Dodiya and Amin, 2015) also exhibited no anthraquinones. Sankhalkar and Vernekar (2016), however, reported the presence of anthraquinone from leaf of *M. oleifera* collected from Chowgule College, Goa (India). They reported more or less similar spectrum of phytochemicals in leaves and flowers. In addition to these phytochemicals, Udofia *et al.* (2020) reported coumarins, lipid and terpenoids in *M. oleifera*. Singh *et al.* (2021) have comprehensively described phytoconstituents of *Moringa* in their review based on 89 references.

Table 1. Physico-chemical characteristic of *M. oleifera* seeds from Kohat, NWFP and Sindh, Pakistan (Anwar and Rashid, 2007) and different localities of Egypt viz. M. Asuit, M. Oraby and M. Monofya (Barkat and Ghazal, 2016).

Phytochemicals (%)	Localities				
	NWFP, Pakistan	SINDH, Pakistan	M. Asuit, Egypt	M. Oraby, Egypt	M. Monofya, Egypt
Seed oil content	34.80	40.39	29.61	30.06	28.62
Moisture	8.90	5.70	7.5	6.57#	6.54
Avail. Carbohydrate	-	-	20.03	19.0	20.29
Fibre	7.54	7.20	10.92	12.16	11.05
Ash	6.53	6.60	4.73	5.06	4.22
Protein	31.65	29.36	35.54	34.51	36.53
Calories value	-	-	450.36	451.1	451.3

The phytochemical composition of *M. oleifera* is reported to vary with the stages of the maturation of the plant after pruning. Early stage (10<sup>th</sup> week after pruning has highest carbohydrate concentration (55.14%), at mid stage (15<sup>th</sup> week) moisture is high (6.3%) and late stage (20<sup>th</sup> week) protein is the highest (28.08%) (Bamishaiye *et al.* (2011) among the stages. The diverse bioactive chemicals are active in several ailments. Anwar and Rashid (2007)

have presented physico-chemical characteristics of seed and seed oil from Kohat, NWFP (Khyber Pakhtunkhwa) and Sindh and Barkat and Ghazal (2016) from some Egyptian localities (Table 1). Seed oil content is significantly high in Sindh plants but protein content in somewhat lesser amount. The oil content of seeds was 34.80% from Kohat and 40.69% from Sindh – quite higher than that in Egyptian accessions. Seeds were rich in protein as well (around 30%) but somewhat higher in Egyptian plants. High oleic acid content (around 73.22%) was reported in local Egyptian provenance (Barkat and Ghazal (2016) who suggested that *Moringa* seed oil may be used for edible and commercial applications. Udofia *et al.* (2020) presented phytochemical composition of leaves as outlined in Table 2. In Kenya sample protein concentration was significantly lower as compared to that of Kohat and Sindh of Pakistan. Protein content of leaves of *Moringa* rivals the protein contents of eggs. Leaves have more protein than seeds (Aleksic 2020). Ash, arachidic acid, brassicasterol,  $\beta$ -carotene, campestanol, carbohydrates, stearic acid, 4-(alpha-L-rhamnosterol)-benzylglucosinolate, 28-isovenasterol, alpha-tocopherol, 4-(alpha-L-rhamnosyloxy) benzyliso Thiocyanate, Behenic acid, Beta-sitosterol, fiber, oleic acid, palmitic acid, proteins, water, 2,4-methylene-cholesterol etc. are but few general phytochemicals of *Moringa* (Misra *et al.*, 2011; Abdulkarim *et al.*, 2005).

Table 2. Phytochemicals of *M. oleifera* leaves from Kenya (Udofia *et al.*, 2020).

Phytochemicals	Leaf - Mean % composition g/100g DW
Carbohydrates	41.97 ± 0.72
Flavonoids	1.12 ± 0.20
Alkaloids	0.06 ± 0.0
Phenols	46.81 ± 3.20
Saponins	14.00 ± 0.51
Ash	14.82 ± 0.65
Crude fibre	1.58 ± 0.45
Crude Lipid	6.06 ± 1.63
Crude Protein	23.05 ± 0.62
Moisture	12.92 ± 0.02

Fahal *et al.* (2018) have reported on phytochemicals of pods of *M. oleifera* – alkaloids 3.1, Flavonoids 5.2, saponins 6.5, sterols 4.7 and tannins 22.9 %, substantially different from Kenyan leaves (cf. Table 2). A number of elemental studies have been conducted by some workers on *M. oleifera* (Udofia *et al.*, 2020 from Kenya; Valdez-Solana *et al.*, 2015 from Mexico; Nweze *et al.*, 2014 from Nigeria, Zaini *et al.* (2019) from Algeria, Leone *et al.* (2015) from Chad and Haiti and Yaméogo *et al.* (2011) from Berkina Foso. There was presence of several heavy metals (Table 3) in differentially varying amounts with locality. Moyo *et al.* (2011) reported Selenium and Boron to be 363 and 49.3 mg /kg DW in *Moringa* leaves. The content of Arsenic in San Pedro (Mexico) sample was reported to be higher than the permissible limit. According to Ganantra-Tejas *et al.* (2012) *Moringa* is richer in Ca (4X of Milk), K (63 X of Milk and 3X of Banana), Mg (36 X of Egg), and Fe (25X of Spinach). They further reported that it has protein (2X of Yoghurt / Milk; Polyphenol (8X of red wine) and Amino acids (2X of Black vinegar. Islam *et al.* (2021) reported Mg to be 635 ± 8.66 mg/100g and P (75 mg / 100g).

Table 3. Elemental analysis of *Moringa oleifera* from various countries. (\*, Higher than the permissible limit.).

Elements (mg / 100 g)	Kenya (Udofia <i>et al.</i> , 2020)	Mexico (Valdez-Solana <i>et al.</i> (2015)		Nigeria (Nweze <i>et al.</i> 2014)-	Southern area of Algeria (Zaini <i>et al.</i> , 2019)	Chad & Haiti (Leone <i>et al.</i> , 2015)	Berkina Faso (Yaméogo <i>et al.</i> , 2011)
		Lombarda	San Pedro				
Cu	0.08 ± 0.07	1.03 ± 0.47	0.41 ± 0.0	10.0	0.81 ± 0.01	ND	NP
Mn	0.84 ± 0.04	-	-	-	5.21 ± 0.03	ND	NP
Fe	0.21 ± 0.13	19.37 ± 6.6	7.07 ± 0.4	30	39.0 ± 3.0	17.03 ± 0.79	19.8 ± 0.0
Zn	6.65 ± 1.51	1.0 ± 0.7	1.06 ± 0.6	5	3.37 ± 0.09	2.48 ± 0.01	2.2 ± 0.0
Ca	1652.3 ± 4.02	-	-	2.090	2785.0 ± 1.0	1839.1 ± 12.3	2100 ± 2.0
Mg	37.07 ± 0.11	322.5 ± 0.0	340.6 ± 2.8	480	382.0 ± 1.0	562.49 ± 9.07	313.0 ± 0.0
Na	163.90 ± 8.09	8.13 ± 0.6	40.78 ± 0.7	-	319.6 ± 6.0	307.65 ± 1.99	ND
K	304.5 ± 57.10	1845.0 ± 7.0	1817 ± 14.5	1620	1626.0 ± 1.0	NP	2250.0 ± 2.0
Ca + Mg	-	2016.5 ± 22.6	2620.5 ± 5.6	-	-	-	-
Se	-	0.00955 ± 0.0	0.107 ± 0.0	-	-	-	-
Pb	-	0.355 ± 0.0	0.20 ± 0.0	-	-	-	-
As	-	0.0055 ± 0.0	0.28 ± 0.0 *	-	-	-	-
S	-	-	-	850	-	-	-
P	-	-	-	40	-	-	-

ND, not determined; NP, Not performed.

Phenolics and flavonoids are the most important antioxidants in *M. oleifera* leaf. Manguro and Lemmen (2007) have described thirteen phenolics from leaves of *M. oleifera* viz. Flavonol glycosides – Kaempferide 3-O – (2'', 3'' diacetylglucoside, Kaempferide 3-O-(2''-O-galloylrhamnoside), Kaempferide 3-O- (2''-galloylrutinoside) -7-O-alpha-rhamnoside, Kaempferol 3-O-[beta-glucosyl-(1 ->2)]-[alpha-rhamnosyl-(1→2)]-[alpha - rhamnosyl -(1 -> 4)]-beta glucoside

-7O-alpha-rhamnoside, Benzoic acid 4-O-alpha glucoside, Benzoic acid 4-O- alpha -rhamnoside – (1 - - > 2) betaglucoside, Benzyldehyde 4-O-beta-glucoside, Kaempferol 3-O-alpha-rhamnoside, Kaempferol, Syringic acid, Gallic acid, Rutin and Quercetin-3-O-beta-glucoside. Phenolics are larger in amount in leaf than that in flowers (Sankhallar and Vernekar (2016). They reported two new flavonoids, Biflavonyl and Myrcetin in flowers of *Moringa*.

Niziot-Tukaszewska *et al.* (2020) have also detected several polyphenols in *M. oleifera* such as Quinic acid, Chlorogenic acid, Gallic acid, Coumaroyl-quinic acid, Kaempferol-3-O-glucoside, Kaempferol-3-O-rutinoside, Rutin, Quercetin-acetyl-glucoside, Quercetin-acetyl-glucoside, Quercetin-malonyltheoxiside, Isoquercetin, Kaempferol-acetyl-glucoside and Quercetin. The plant provides a rich and rare combination of Zeatin, Quercetin,  $\beta$ -Sitosterol and Kaempferol (Biswas *et al.*, 2012). Numerous more phytochemicals may be seen in publications of Foidl *et al.* (2001), Yongbi (2010), Oluduroa *et al.* (2010), Amaglo *et al.* (2010), Sharma *et al.* (2011), and Bhalla *et al.* (2021).

Okumu *et al.* (2016) described phenolic content of aqueous and aqueous + methanol extracts of *M. oleifera* to be  $34.42 \pm 5.80$  and  $52.04 \pm 3.12$  mg of Gallic acid equivalent per g of the dry plant material (mg GAE.g<sup>-1</sup>), respectively, in Kenyan accession of *M. oleifera*. The total flavonoids content was  $79.13 \pm 13.04$  and  $366.09 \pm 89.96$  mg of Catechin equivalent per g plant material (mgCE.g<sup>-1</sup>), respectively, in this Kenyan accession (Okumu *et al.* (2016). In this accession, ascorbic acid content was  $2.02 \pm 0.66$  and  $3.04 \pm 2.06$  mg of ascorbic acid equivalents per g dry plant material (mg.AAE.g<sup>-1</sup>) when extracted in water and water + methanol. In South Indian accession of *M. oleifera* total phenol and total flavonoids were  $627 \pm 12.26$  mg Gallic acid equivalent / 100g and  $22.16 \pm 1.54$  mg Quercetin equivalent /g, respectively (Shanmugavel *et al.*, 2018).

Table 4. Amino acids contents in leaf and seed of *Moringa* (Source: El-Shohaimy *et al.*, 2015; Ijarotimi *et al.*, 2013).

S. No.	Amino acids	El-Shohaimy <i>et al.</i> (2015)	Ijarotimi <i>et al.</i> (2013)	Seed / leaf ratio
		Leaf (mg / 100g)	Seeds (mg/100g)	
1.	Lysine *	63.13	312	4.93
2.	Histidine *	29.56	1930	66.4
3.	Valine *	62.34	1080	17.32
4.	Leucine *	94.36	3830	40.59
5.	Isoleucine *	46.98	4230	90.04
6.	Threonine *	48.35	3020	62.46
7.	Alanine	4.93	5160	1046.7
8.	Aspartic acid	13.76	1570	114.10
9.	Serine	3.13	3060	977.6
10.	Proline	1.86	2180	1172.04
11.	Glutamic acid	18.03	17870	991.1
12.	Glycine	2.31	2370	1025.97
13.	Arginine *	7.65	8280	1082.4
14.	Cysteine o	2.15	1680	781.4
15.	Tyrosine	2.03	1970	970.4
16.	Methionine*, o	0.43	310	720.9
17.	Phenylalanine	3.42	3270	956

\*, Essential amino acids. o, Sulphur containing aminoacids.

The phytochemicals of medicinal interest in this species include flavonoids, glucosinolates and phenolic compounds, tannins, saponins, alkaloids, anthraquinones, glycosides, steroids etc. (Saleem, 1995; Manguro and Lemmen, 2007; Kasolo *et al.*, 2010; Amaglo *et al.*, 2010; Idris and Adamu, 2018) which may vary in concentration depending on the climatic conditions, the method of collection and processing (Coppin, 2008; Mukunzi *et al.*, 2011). In addition, other compounds have been isolated from leaves, including different types of glycosides (Faizi and coworkers, 1998, 1997, 1995, 1994, Murakami *et al.*, 1998; Bennett *et al.*, 2003). Leaves also contain carotenoids, tocopherols and vitamins. The root bark is rich in two alkaloids: moringine and moringinine. It may, however, be mentioned here that very important compounds such as Niazirin, Niazirin, and three mustard oil glycosides 4-(4'-O-acetyl-alpha-L-rhamnosyloxy) benzyl isothiocyanate and Niaziminin A and Niaziminin B were first isolated by Faizi *et al.* (1994) from ethanolic extract of *Moringa* leaves. It was the first report of isolation of nitriles, and isothiocyanate and isothiocarbamates from the same plant. Isothiocyanate and thiocarbamate glycosides Niaziminin A and B showed hypotensive activity while nitrile glycosides were found to be inactive in this regard. Other important publications on phytochemicals of *M. oleifera* were by Faizi *et al.* (1997) regarding isolation and

El-Shohaimy *et al.* (2015) and Ijarotimi *et al.* (2013)) have reported amino acids contents in leaves and seeds of *M. oleifera*. Seed / leaf ratio of various amino acids as calculated from their data is presented in Table 4. Obviously seeds were richer in amino acids contents by many folds – more so in case of Alanine, Serine, Proline, Glutamic acid, Glycine, Arginine and Tyrosine. A comparative account of essential and non-essential amino acids in eggs, chicken breast, Ruifish, Pongas fish, Tilapa fish, soybean, rice, and wheat flour was made by Islam *et al.* (2021). *M. oleifera* profile was richer than many animal- and plant-derived sources, and more or less similar to that of egg and chicken breast. That's why it is reported to improve muscle strength and detoxify body (Basillo-Heradia and Gutierrez-Grijalava, 2022).

structural elucidation of Niazidin (a novel glycoside) from pods and Faizi *et al.* (1998) regarding hypotensive constituents of *Moringa* pods.

Compounds of *M. oleifera* such as 4-(4'-O-acetyl- $\alpha$ -L-rhamnopyranosyloxy) benzyl isothiocyanate, 4-( $\alpha$ -L-rhamnopyranosyloxy) benzyl isothiocyanate, Niazimicin, Pterygospermin, Benzylisothiocyanate and 4-( $\alpha$ -L-rhamnopyranosyloxy) benzyl glucosinolate (Glucomoringin), Benzyl isothiocyanate 4-(-L-rhamnopyranosyloxy), Aglycone of Deoxy-Niaziicine (N-benzyl, S-ethyltioformate), 4-hydroxymullein, Moryngyne, Niaziridin, Niazirin, and Isothiocyanatomethyl benzene (Fahey, 2005; Singh *et al.*, 2017; Pandey *et al.*, 2019) are unique and responsible for the properties of this species (Singh *et al.*, 2017; Pandey *et al.*, 2019). Glucomoringin is the most abundant glucosinolates in *Moringa* to be as high as > 30mg/g DW in seeds and 20 mg/g DW in leaves (Baeza-Liménez *et al.*, 2022). The two alkaloids – glomoringin and glucosoonjnain are reported to vary by Chodur *et al.* (2018) in domesticated and wild types as evaluated in 36 accessions (21 domesticated and 15 wild types) collected from Pakistan, Madagascar, Kenya, Thailand, India, USA, Germany and South Africa. The wild types were organoleptically bitter than domesticated types. There was substantially higher average concentration (75.29  $\mu$ mol/g DW) of glucomoringin in domestic types than in wild types (18  $\mu$ mol/g). On the other hand, average glucosoonjnain concentration (33.79  $\mu$ mol/g) was significantly higher in wild types as compared to 1.6  $\mu$ mol/g in domesticated types (Chodur *et al.*, 2018).

Karthika *et al.* (2013) have isolated some 28 compounds from chloroform extract of *M. oleifera* leaves of which several were reported to be useful one way or the other as given below:

Isopropyl alcohol (antibacterial, antiseptic, antiesthatic, surfactant and Neurolytic), Ethyl alcohol (Antibacterial, antiseptic, antiesthatic, surfactant), Propylene glycol (Used in pharmaceutical), Lactic acid (Antibacterial agent, Food additive), Ethyl 2-hydroxypropanate (lactate) (Food additive), Phenethyl alcohol (antibacterial), Pyrocatechol (Pesticide, flavours, fragrances), 2, 3-Butanedione (DAS) Diacetyl (Flavours, Fragrances, alcoholic beverages), Linalool oxide (Antioxidant, antimicrobial), Trans-Linaloloxide (Antioxidant, antimicrobial), Heptanol (Cosmetics for fragrance), 1,2-benzenedicarboxylic acid, diethyl ester (CAS) ethyl phthalate (Plasticizer, antibacterial), Adenine (Antiviral, diuretic, antianemic), Palmitic acid (Antioxidant, pesticide, antimicrobial, lubricant) and 1,2-benzenedicarboxylic acid, bis (2-ethyl hexyl) ester (antibacterial).

Some 163 phytochemicals have been reported in *Moringa* by Lui *et al.* (2021) recently. Some of these compounds are unique to the *Moringa* family.

## 2) Vitamins contents of *M. oleifera*

The plant is rich in vitamins and minerals (Dahot, 1988) - even richer than several well-known sources as given below (Ganatra-Tejas *et al.*, 2012). This makes *Moringa* a valuable nutritional plant. Its edible parts have:

Vitamin A – 4X of carrot and 13 x of Spinach. Fresh leaves contain Vitamin A around 6.8 mg per and dried leaves contain 18.9 mg per 100g dried leaves (Hanuk, 2018).

Vitamin B – 4 X of meat; rich in Vitamin B2 (Riboflavin); Vitamin B3 (Niacin) – 50 x of Peanut; Vitamin C – 7X of oranges. Hanuk (2018) reported 220 mg of Vitamin C in 100g fresh leaves but only 17.3 mg in dried leaves of *Moringa*. Adenike (2014), however reported that sun-dried samples have higher contents of Vitamins C and E while samples dried at ambient conditions have higher concentration of P, K, Mg, and Vitamin A. Adenike (2014) thus concluded that *Moringa* leaves should be dried at ambient conditions due to higher retention of some essential minerals and Vitamins. Seeds are highly rich source of Vitamin E (751  $\pm$  4.41 mg/100g plant material) as reported by Islam *et al.* (2021).

## 3) *M. oleifera* in human feeding

James and Zikankuba (2017) have described this plant as potential tree for nutrition security in Africa. NASA included it in list for space food. Isitua *et al.* (2015) advocated the use of *Moringa* leaves as food. Leaves have no trans fatty acids. The leaves of *M. oleifera* are used in preparing soup, salad and for making tea and other culinary uses (Stevens *et al.*, 2013). It is rated as super food (<http://pharmaesy.in/blog/category/home-remedies/>). It is used in soup, chocolate, biscuits and cakes, bread (adding 5% leaf powder in flour) and muffin (12% leaf powder) and several other food preparations (Islam *et al.*, 2021). Leaf-based food fights against malnutrition.

One year old *Moringa* tree is able to produce flowers and pods. Flowers are rich in Ca and K. Flowers may be mixed with batter and deep fried and eaten as highly nutritious special food (Halder and Kosankar, 2007). The cooked flowers have a taste like mushrooms. The flower nectar gives honey (Misra *et al.*, 2011).

The green seeds may be peeled off from outer coating and may be eaten as pea. In Malaysia, the green pods are used as ingredients of local curry varieties. From the roots sauces are prepared. The leaves show high content of vitamins, pro-vitamins and minerals (Palada and Chang, 2003). They also contain all essential amino acids including arginine and histidine, which are generally found in proteins of animal origin. The amino acids are very important

for children's growth. FAO promoted a program for the use of *M. oleifera* aimed at the children's population at malnutrition and the pregnant and lactating mothers (Fuglie, 2001). The *M. oleifera* oil is rich in oleic acid and tocopherols (Anwar *et al.*, 2005). Except for its lower content of linoleic acid, this oil shows similar chemical composition and physical properties as olive oil. It is used in salad seasoning in Haiti and other Caribbean islands (Foidl *et al.*, 2001) with no adverse effects, allergies or toxicity (Ghazali and Mohammed, 2011).

The oil cake of *Moringa* seeds is bitter in taste and known to contain anti-nutritional elements like haemagglutinins, glucosinolates, alkaloids and saponins (Farooq and Rashid, 2007).

The alkaloid moringinine in root bark is toxic and may cause nervous problems. Pterygospermin and alkaloid spirachin are nerve paralyzing (Halder and Kosankar (2007) and rated to be antibacterial (Das *et al.*, 1957). Addition of Moringa extract in butter at the rate of 800 ppm caused a decrease in product acceptance significantly as Moringa altered the organoleptic properties (Nadeem *et al.*, 2013). Seed oil addition in ice cream was not effective in preventing the formation of free fatty acids (Nadeem *et al.*, 2016).

The composition of leaf flour of Moringa, its digestibility and antinutrition contents are presented in Table 5A as reported by Teixeira *et al.* (2014). The leaf flour has low contents of the antinutritional substances). Ferreira *et al.* (2008), also reported low amounts of tannins (12 mg g<sup>-1</sup>) and absence of cyanogenic compounds in *M. oleifera* leaves. The content of oxalic acid was reported by Teixeira *et al.* (2014) to be much lower (10.5 mg/g) (Table 5A) than that in spinach (822 mg) by Franco (2005). Oxalic acid was 1.35 ± 0.03 g/ 100g DW of leaves of Moringa in Algerian accession (Zaini *et al.*, 2019). The trypsin inhibitors in Moringa were found much less active (Teixeira *et al.*, 2014) than those in soybean (107.22 trypsin Inhibitors units.mg<sup>-1</sup>) as reported by Da Silva and Kerr (1999). Teixeira *et al.* (2014) compared their results on antinutritional substances to that in taro (reported by Pinto *et al.* (2001) and concluded that *M. oleifera* leaves can be eaten without nutritional damage. However, cyanogenic glucosides have been reported from Nigerian *M. oleifera* to be 3.3% in leaves and 2.60% in flowers (Okah and Carnelius, 2019) (Table 5B). The production of cyanogenic compounds appear to be environmentally regulated.

Table 5A. Composition of leaf flour, its in vitro digestibility and antinutrition substances (data from Teixeira *et al.*, 2014). Trees were located in Minas Gerais, SE Brazil).

S. No.	Substance	% concentration
1.	Crude Protein	28.7 *
2.	Fat	7.1
3.	Ashes	10.9
4.	Carbohydrate	44.4
5.	Ca	3.0 mg / 100g
6.	Fe	103.1 mg / g
In vitro digestibility (%)		
1.	Casein (standard)	100
2.	Defatted flour	33.29
Antinutrition substances		
1.	Total tannins	20.60 mg/g
2.	Trypsin inhibitor	1.45 TUI/g
3.	Nitrate	17.0 mg/g
4.	Oxalic acid	10.5 mg /g
5.	Cyanogenic compounds	Absent
Others		
1.	B- carotene	161.0 µg/g
2.	Lutein	47.0 µg/g

\*, Teixeira *et al.* (2014) opined that protein although in substantial amounts, but of low digestibility which suggests that *in vivo* studies should be carried out to assess their use.

#### 4) Pharmaceutical and medicinal properties

Pharmacologically, Moringa is antioxidant, antidiabetic, anti-spasmodic, diuretic, ecboic, anticancer, anti-microbial, anti-inflammatory, anti-hyperlipidaemic, anti-hyperglycemic, hepatoprotective, antiasthma, anthelmintic, anti-fertility, antipyretic, analgesic, anticonvulsant, renal protector, hypocholesteromic, anti-asthmatic, etc. (Das *et al.*, 1957; Shaw and Jana, 1982; Tahir *et al.*, 2017; Prithiviraj and Sumathy, 2021). And considered "Maha Oshidhi" (great medicine) in Ayurvedic system of medicine. It treats diseases (respiratory, gastrointestinal, inflammatory, cardiac, nutritional and skin diseases (Velazquez-Zavala *et al.*, 2016; Agarwal and Mehta, 2008; Mahajan *et al.*, 2009). Razis *et al.* (2014) have described it as antitumour, anticancer and antifibrotic also. It is of potential benefit in Lactating mothers, menopause, depression, osteoporosis, rather functional in more than eighty diseases

Table 5B. Phytochemicals in leaves and flowers of Nigerian Moringa (Okah and Carnelius, 2019).

	Leaves (%)	Flowers (%)
Saponins =	5.00	3.20
Flavonoids =	5.42	7.12
Alkaloids =	5.36	1.55
Cyanogenic		
Glycosides =	3.30	2.60

Okah and Carnelius (2019) reported that saponins and flavonoids may be helpful in reducing blood cholesterol and prevention of cancer as suggested by some authors. The presence of cyanogenic glycosides should produce hydrogen cyanide in body but that could be easily detoxified by the body. This, obviously, needs to be studied in detail.

Moreover, Ojiako (2014) reported tannins to be 8.22% in leaves of Moringa of Anambra State, Nigeria- in quite higher concentration than that of Brazil plants (Teixeira *et al.*, 2014). This may probably be due to very much arid conditions of Nigeria

(Mahmood *et al.*, 2010). Besides other uses, it is described as remedy of cholera, conjunctivitis, eye and ear infections, hysteria, Psoriasis, leukoderma, biliousness, semen deficiency, etc. (Das *et al.*, 1957; Shaw and Jana, 1982; Shamim *et al.*, 2018). The traditional and pharmacological benefits of *M. oleifera* plant, described in many publications (to cite a few, Delaveau and Baiteau, 1980; Ramachandran *et al.*, 1980; Villasenor *et al.*, 1989 a and b; Makkar and Backer, 1996, 1997, 1999; Njoku and Adikwu, 1997; Fuglie, 1999; Rao *et al.*, 2001; Fahey, 2005; Agbogidi and Ilondu, 2012; Stevens *et al.*, 2013; Ganatra-Tejas *et al.*, 2012; Shamim *et al.*, 2018; Basillo-Heradia and Gutierrez-Grijalava, 2022 and several others), may be briefly summarized as follows:

**Roots:** Roots are reported to be antilithic, rubefacient, vesicant, carminative, antifertility, anti-inflammatory, stimulant in paralytic afflictions, acts as a cardiac / circulatory tonic, treating rheumatism, inflammations, articular pains etc. (Swati *et al.*, 2018). **Leaves:** Agbogidi and Ilondu (2012) described 75 therapeutic and prophylactic uses of *M. oleifera* from various sources. Medical use of leaf in Nigeria included curing of fever, treatment of ear infections and blood pressure (Stevens *et al.*, 2013). Swati *et al.* (2018) have reported application of its leaf poultice to sores, on temples for headache and in piles etc. **Flowers:** Throat infection, common cold, external sores, Anthelmintic, Antitumor, Rheumatism, tonic, Diuretic, hysteria and Abortion (Makkar and Becker, 1996, 1997, 1999; Ganatra-Tejas *et al.*, 2012). Flowers of *M. oleifera* are reported to be cholagogue, tonic, anti-oxidant, prescribed in cold phlegmatic condition (Rizvi, 1998; Rizvi and Ali, 2016) and epilepsy (Fuglie, 1999). **Seeds (Nuts):** Ganatra-Tejas *et al.*, (2012) described seed to be anthelmintic, useful in warts, Anti-tumor, Ulcers, rheumatism, arthritis, antispasmodic, goitrogen, and minerals / Vitamins deficiency. Seeds of *Moringa* are known to contain Vitamin A. *Moringa* juice is useful in conjunctivitis (Rao *et al.*, 1999). Seed extract of *M. oleifera* may be used as disinfectant (Bichi *et al.*, 1995). Seed extract of *M. oleifera* is antimicrobial (Eilert *et al.*, 1981). Frying quality of seed oil is said to be better in *M. oleifera* as compared to other vegetable oils (Abdulkarim *et al.*, 2007). Seeds are also used in perfume industry, cosmetics, lubricant, soap, oil and as body cream (Bhargave *et al.*, 2015). It is useful in pyoderma. It is anti-spasmodic, anti-schistosomes and antibacterial. **Fruits:** Anthelmintic, Skin Cancer, Anti-hypertensive, Diabetes, Cardio tonic, Joint Pain and drunculiasis. **Seed oil:** Bladder disorders, hepatomegaly, prostrate function, antioxidant, purgative, dermal application, fungal /mycoses. **Exudates:** Dental Caries, Toothache, Syphilis, Typhoid, Earache, Fever, Asthma, Diuretic, Dysentery, Rheumatism, Headache, Abortifacient, Rubrafacient. **In Vitro:** Antibacterial and anti-fungal activity – *Bacillus cereus*, *B. subtilis*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Streptococcus faecalis*, *S. aureus*, *S. epidermidis*, *Shigella shinga*, *S. sonnei*, *Aspergillus niger*, *Candida albicans*, *Microsporum canis*, *Fusarium solani*, *Rhizopus solani*, *Yeast dermatophytes* and *Epidemophyton xoccosum*. *M. oleifera* is active against helminths – *Trichophyton rubrum*, *T. mentagraphytes*. Compounds like 4-(-L-rhamopyanosyloxy) benzyl iso-thiocyanate, 4- (-L- rhamopyanoxyloxy) benzyl-glucosinolate and Pterygospermin are considered to be responsible for antibiotic activity.

Table 6. Antioxidant activity of *M. oleifera* on the basis of various antioxidant parameters.

Antioxidant Parameters	Methanol-Water Extract	Hydrophilic Extract	Lipophilic Extract
TEAC ( $\mu\text{mol TE}/ 100\text{g FW leaves}$ )	6208 $\pm$ 206	1426 $\pm$ 72	341 $\pm$ 16
DPPH ( $\mu\text{mol TE}/ 100\text{g FW leaves}$ )	3055 $\pm$ 140	854 $\pm$ 64	125 $\pm$ 10
FRAP ( $\mu\text{mol TE}/ 100\text{g FW leaves}$ )	3962 $\pm$ 297	635 $\pm$ 22	580 $\pm$ 50
ORAC ( $\mu\text{mol TE}/ 100\text{g FW leaves}$ )	10805 $\pm$ 690	6683 $\pm$ 204	201 $\pm$ 11
TPC (mg GAE / 100g FW leaves)	504 $\pm$ 28	145 $\pm$ 6	63.5 $\pm$ 4.9
Tobart index	0.2058	0.2423	0.2416

Source: Gonzalez-Romero *et al.* (2020). Abbreviations: TEAC = Trolox equivalent antioxidant capacity; DPPH = 2, 2-diphenyl - 1-picril-hydrazyl hydrate; FRAP = Ferric ion reducing antioxidant Power; ORAC = Oxygen radical Absorbance capacity; TPC = Total Polyphenol content by Folin Ciocalteu reagent. TE = Trolox equivalent per 100g FW. In each case the *M. oleifera* activity was the highest.

### 5) Antioxidant property

*Moringa* is highly rich in antioxidants, a most probable reason of its health benefits. The flavonoids act as antioxidants and enhance the effects of vitamin C (Korkina and Afanas'ev, 1997). The Flavonoids are also effective antimicrobial and have the ability to bind with extra-cellular and soluble proteins and complexes of bacterial cell walls. Phenolics and flavonoids can form chelate complexes with metal ions, thereby getting easily oxidized by donating electron to scavenge free radicals (Siddhuraju and Becker, 2003; Sreedam *et al.*, 2010). Higher phenolic concentration in *M. oleifera* is correlated with increased antioxidant activity of *Moringa* (Kostyuk *et al.*, 2001; Borneo *et al.* (2008).

*M. oleifera* possess highly potent antioxidants in leaves and seeds (Unuigbe *et al.*, 2014; Niziot-Tukaszewska *et al.*, 2020). According to Pakade *et al.* (2013) *M. oleifera* contains more than 40 natural antioxidants and well known for its effects in elimination of free radicals. Gonzalez-Romero *et al.* (2020) have investigated antioxidant potential

of *M. oleifera* grown in Andalusia (Spain) and compared it with 28 types of salad plant or their leaves (green lettuce, red lettuce, beet leaves, chand, Chinese cabbage, curly green cabbage green leaves, curly green cabbage white leaves, endive, escarole, green cabbage, Green lettuce plants, iceberg lettuce plants, kale, lamb's lettuce, lollo rosso lettuce (mix salad (white cress + lamb's lettuce + lollo rosso salad), Oak leaf lettuce, red Cabbage, red chickory, red lettuce plants, red lettuce sprouts, rocket salad, roman rettuce salad, spinach, spinach sprouts, turnip green (tops) and water cress) on the basis of five antioxidant parameters. It was found to be the best antioxidant plant always at the first rank in antioxidant efficiency – much higher in activity than the test species. The antioxidant activity of *M. oleifera* is given in Table 6. The methanol-water extract showed the maximum activity. The leaves were rich in total phenolic content, carotenoids, total flavonoids and chlorophylls. It was concluded by Gonzalez-Romero *et al.* (2020) that addition of *M. oleifera* leaves to the existing range of fresh-cut salad foods would increase their antioxidant content by six times. Sajid *et al.* (2020) showed that seed oil of *M. oleifera* is antioxidative hypoglycemic agent capable of improving the other clinical conditions as well related to oxidative stress of diabetes mellitus such as hepatic, renal and pancreatic functions in alloxan treated rats. Luqman *et al.* (2012) have also shown the leaf and fruit extracts of *Moringa* to be potent antioxidant. In a sample from Mardan (Pakistan), Iqbal and Bhangar (2006) had analyzed samples from Balakot, Chakwal, Jamshoro, Nawabshah and Mardan and found that the antioxidants content of *M. oleifera* was higher in the month of December or March and least in Jume depending upon the location i.e., seasons and agro-climatic location influence antioxidant activity. Atawodi *et al.* (2010) suggested the plant's high polyphenol content to be related to antioxidant activity. Sidhuraju and Becker (2003) also found variation in antioxidant properties of *M. oleifera* with solvent extraction of phenolic constituents from three different agro-climatic origins. According to Olaoye *et al.* (2021) environmental parameters such as annual precipitation, minimum and maximum temperatures and soil type influence the antioxidant activity of *M. oleifera*. The flavonoids, total phenolic content, saponins, lipid peroxidation inhibition, tannins contents varied with samples locations (21 accessions) in Southwestern States of Nigeria. Percent antioxidant activity as DPPH scavenging activity and nitric oxide scavenging activity was directly related with the dose (positive dependency). Some very important antioxidants in *Moringa* are investigated and discussed in Bennet *et al.* (2003), Turner *et al.* (2015), Amara *et al.* (2021) etc. Most of the health benefits of *Moringa* appear to be due to its antioxidants and their richness.

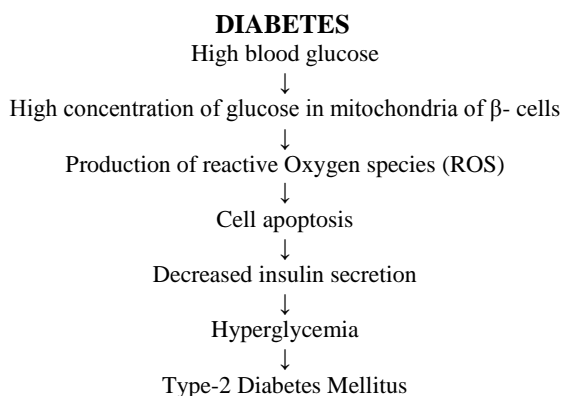


Fig. 1. Metabolic sequences of high glucose leading to Type-2 Diabetes in man. *Moringa oleifera* prevents the apoptosis of  $\beta$  – cells due to its antioxidant contents. Which combine with the ROS and prevent the cell damage. Fig. source: Gopalakrishnan *et al.* (2016).

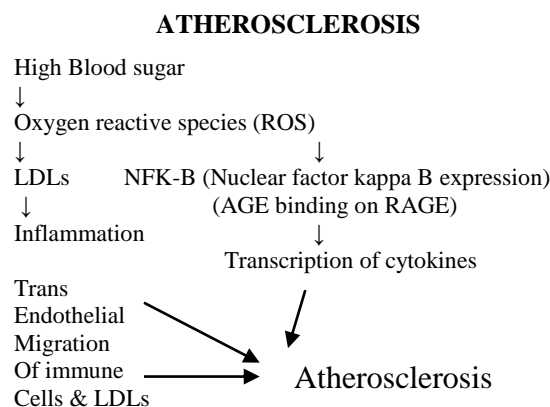


Fig. 2. Diabetes leading to atherosclerosis. Abbreviations: AGE, Advanced glycelated end products; RAGE, Receptors for AGEs on surface of cells. *Moringa* can prevent atherosclerosis by scavenging ROS and preventing formation of AGE and LDLs. Seen in Gopalakrishnan *et al.* (2016).

Antidiabetic effects of *Moringa* have been reported in literature (Kaneto *et al.*, 1999; Mbikey, 2012; Cerf, 2013; Al-Malki and El Rabey, 2015; Villarreal-Lopez *et al.*, 2018). There were significant hypoglycemic effects of *Moringa* in diabetic rats and no lethal dose was discovered. Isothiocynates rich extract of foliage is reported to act against insulin resistance (Waterman *et al.*, 2015). Chumark *et al.* (2008) have discussed anti-atherosclerotic effects of *M. oleifera*. Ezeigbo *et al.* (2016) have demonstrated more hypoglycemic effect of aqueous extract of leaves of *M. oleifera* than its ethanol extract, after 14 days of administration in alloxan-induced diabetic Wistar rats. There was 45.2 and 33.7% reduction in blood glucose for aqueous and ethanol extract, respectively whereas the reference drug insulin had 58.7% reduction. Diabetic and atherosclerotic pathways in diabetics have been elaborately discussed by Gopalakrishnan *et al.* (2016) – (Fig. 1 and 2). *Moringa* prevents diabetes by scavenging ROS and thus



prevention of apoptosis of  $\beta$ -cells due to its anti-oxidative contents. Similarly, *Moringa* can prevent atherosclerosis by scavenging ROS and preventing formation of LDLs and advanced glycelated end products (Aronson and Rayfiels, 2002; Wright *et al.*, 2006; Chumark *et al.*, 2008; Mbikey, 2012; Gopalakrishnan *et al.*, 2016). Rutin, a phenolic of *M. oleifera* is reported to be antihyperglycaemic and antioxidant (Kamakkannan and Prince, 2006).

Table 7. List of susceptible organisms to *Moringa oleifera*. (Source: Xiao *et al.*, 2020).

S. No.	Organisms	Active Plant component
Bacteria		
1.	<i>Aeromonas cavi</i>	Leaf
2.	<i>Bacillus cereus</i>	Seed
3.	<i>B. megaterium</i>	Bark
4.	<i>B. subtilis</i>	Leaf
5.	<i>Citrobacter freundii</i>	Bark
6.	<i>Enterobacter aerogenes</i>	Leaf and seed
7.	<i>Enterococcus faecalis</i>	Leaf
8.	<i>Escherichia coli</i>	Leaf, seeds and pods
9.	<i>Klebsiella pneumonia</i>	Leaf
10.	<i>Mycobacterium phlei</i>	Leaf
11.	<i>Proteus mirabilis</i>	Seed
12.	<i>Providencia stuartii</i>	Leaf
13.	<i>Pseudomonas aeruginosa</i>	Leaf and seed
14.	<i>Salmonella typhimurium</i>	Pod
15.	<i>Serratia marcescens</i>	Leaf and seed
16.	<i>Staphylococcus aureus</i>	Leaf, seed, pods and bark
17.	<i>S. epidermidis</i>	Pods
18.	<i>S. pyogenes</i>	Leaf and seeds
19.	<i>Vibrio cholerae</i>	Leaf and flowers
20.	<i>V. mimicus</i>	Flowers
21.	<i>V. vulnificus</i>	Flowers
22.	<i>Yersinia enterocolitica</i>	Seeds
Fungi		
1.	<i>Aspergillus flavus</i>	Leaf and seeds
2.	<i>Basidiobolus haptosporus</i>	Leaf
3.	<i>Cryptococcus neoformans</i>	Seeds
4.	<i>Epidemophyton floccosum</i>	Leaf and seeds
5.	<i>Microsporium canis</i>	Leaf and seeds
6.	<i>Penicellium aurantiogriseum</i>	Leaf
7.	<i>Trichophyton mentagrophytes</i>	Leaf and seeds
Viruses		
1.	Epstein- bar virus	Leaf and seeds
2.	Equine herpes virus	Leaf
3.	Foot and Mouth disease	Leaf
4.	Hepatitis virus	Dry powder
5.	Herpes simplex virus	Leaf
6.	Human immunodeficiency virus	Leaf
7.	Infectious bursal disease	Leaf
8.	Rhinovirus	Leaf
Parasites		
1.	<i>Brugia malayi</i>	Gum
2.	<i>Cryptosporidium parum</i>	Leaf
3.	<i>Dracuaculiasis</i>	Leaf
4.	<i>Haemonchus</i>	Seeds
5.	<i>Helminths</i>	Seeds
6.	<i>Hymenolepis nana</i>	Leaf
7.	<i>Leishmania donovani</i>	Flowers
8.	<i>Plasmodium</i>	Seeds
9.	<i>Schistosomes</i>	Flowers
10.	<i>Trypanosoma brucei</i>	Aerial parts

## 7) Antimicrobial activity

Aqueous extract of *M. oleifera* seeds have substantial potential against pathogenic microscopic organisms e.g., *Staphylococcus aureus*, *Bacillus subtilis*, *Escherichia coli* and *Pseudomonas aeruginosa* in dose dependent manner (Saadabi and Abu, 2011; Garga *et al.*, 2019). Aqueous extract was exceptionally inhibitory to *Mycobacterium phlei* and *B. subtilis* (Eilert *et al.*, 1981). *Moringa* seed oil has antifungal activity against *Rhizopus solani* and *Fusarium solani* (Chuang *et al.*, 2007). Ayirezang *et al.* (2020) reported that at 100 mg/ mL of methanol extract, the leaves gave wider zone of mycelia growth inhibition against *Aspergillus flavus* and *A. niger* (Food spoilers) – more than the seed extract. It may be mentioned that the activity was lower than that of sodium benzoate. The minimal inhibition concentration of the extracts was more active at 25 mg / mL. Issa *et al.* (2021) have reported ethanolic extract to be inhibitory and *S. aureus* to be the most sensitive to aqueous and ethanolic extract. N-hexane extract exerted no activity. MIC of Aq. Extract was 60mg/mL for *S. aureus*, 80 mg/mL for *P. aeruginosa* and *E. coli*. The MIC of ethanol extract for 60 mg/mL for *S. aureus*, 80 mg/mL for *E. coli* and 100 mg/mL for *P. Aeruginosa* as indicated through agar well diffusion method. *Moringa* leaf extract compared favourably to nystatin, streptomycin and gentamicin (Oladeji *et al.*, 2020). It showed potency against *Klebsiella* sp., *P. aeruginosa*, *Trichoderma* sp., *Aspergillus flavus*, *Bacillus cereus*, *S. pneumoniae*, *Candida* sp. and *E. coli*. Antibacterial activity of *Moringa* leaf against *S. aureus* and *E. coli* is also reported by Malhotra and Mondal (2018) in their *in vitro* studies and speculated it due to tannins, flavonoids, glycosides, terpenoids, phenols, etc. Abalaka *et al.* (2011) reported that chloroform extract showed remarkable activity against *E. coli*, *Salmonella typhi* and *P. aeruginosa*. Ethanolic and methanolic extracts of *M. oleifera*, collected from salt range of Thal desert of Pakistan and dried under shade, were found to be effective against *Aspergillus fumigatus*, *A. niger* and *Candida albicans*. Antibacterial effects of *M. oleifera* on *E. coli*, *Streptococcus pneumoniae* and *Staphylococcus aureus* have been reported by Idris and Abubakar (2016) from Nigeria.

Xiao *et al.* (2020), based on several references, have presented a long list of susceptible organisms (bacteria, fungi, viruses and parasites) as given in Table 7. Crude ethanolic extract from flowers and callus of *M. oleifera* have also exhibited antibacterial activity (Talreja, 2010).

## 8) Neuroprotective effects

In rat model, Moringa has been shown to have a neuro-protective effects against brain damage and decline in memory (Igado and Olapade, 2016). The data of Amara *et al.* (2021) showed that *Moringa* extract prevents oxidative damage by lowering reactive oxygen species (ROS) formation, restoring mitochondrial respiratory chain complex activities, and, in addition, by modulating the expression of vitagenes. *Moringa* extract also protected mitochondria from apoptosis. Alqahtani and Abasher (2021) reported the potential neuroprotective effect of *M. oleifera* extract against lead-induced brain toxicity. The extract stimulated detoxifying enzymes. Ganguly and Guha (2008) have reported *M. oleifera* to protect against Alzheimer's disease by altering brain monoamine levels and electrical activity. Mechanisms triggering neurodegeneration is unknown but has largely been suggested due to oxidative stress (Calabrese *et al.*, 2007). Moringa acts against neurodegeneration probably due to its polyphenolic contents and other antioxidants (Luqman *et al.*, 2012).

### 9) Nematicidal activity

The extracts of leaves and seeds of *M. oleifera* were reported by Oluwatayo *et al.* (2019) to decrease *Meloidogyne incognata* population, reduce root galling as well as the nematode reproduction resulting in the yield of tomato varieties (UC28B, Roma VF and Riogrande). Leaves extract used at 5% concentration at the rate of 40 mL per pot significantly reduced *M. incognata* population in association of sugar beet CV. Gazelle (El-Nagdi and Youssef, 2015). Nematicidal and antifungal activity of leaves and seeds extracts was also reported by Olajide *et al.* (2018) when used in cucumber (*Cucumis sativus*), *M. incognita* and soil borne fungi such as *Phytophthora* sp., *Rhizopus* sp. *Penicillium* sp. *Aspergillus niger*, *A. parasiticus* and *A. flavus*. All fungi were *in vitro* differentially inhibited by seeds and leaves extract, but Seed extract was inhibitor on *Phytophthora* sp. and leaf extract was inhibitor on *A. flavus*. A variety of cucumber "marketmore" was higher in performance of vegetative growth than variety cucumber Roma-vf. Cold pressed seed oil of *M. oleifera* was reported to be effective against *M. incognata*. Compared to free eggs, egg masses were more sensitive to the oil as the second stage juveniles.

Both alone or combined application of *Trichoderma harzianum* and *M. oleifera* has been reported to cause significant reduction of hatching of eggs and caused higher juvenile mortality of *Meloidogyne javanica* associated with eggplant under greenhouse conditions. Treatment against *Meloidogyne javanica* with *T. harzianum* combined with *M. oleifera* was more effective both *in vitro* and under field conditions (Murslain *et al.*, 2014). A possible mechanism of effectiveness of plant extracts or seed oil is considered to involve disruption of cell membrane of the nematodes (Kayani *et al.*, 2012). The studies are, however, not conclusive (Refaat *et al.* (2020) who opined that there is a need for further research to elucidate the nematicidal activity of seed oil under greenhouse conditions and moreover the physiological mechanism in action.

### 10) Immunity against viruses

Kurokawa *et al.* (2016) reported that aqueous extract of foliage builds immunity against herpes and simple virus type 1 (HSV 1).

### 11) Moringa as breast-milk augment

*M. oleifera* is reported to augment breast milk in lactating women (Singh *et al.* (2021).

### 12) Antitumour and anticancer property

Moringa leaves have been reported to show antitumor activity and niazimicin has been proposed to be a potent chemo preventive agent in chemical carcinogenesis (Guevara *et al.*, 1999). The seed extracts have also been found to be effective on hepatic carcinogen metabolizing enzymes. Bharali *et al.* (2003) have reported that a thiocarbamate from the leaves of *M. oleifera*, exhibited inhibition of tumor promoter induced Epstein-Barr virus activation. Antitumor and anticancer properties of Moringa have been reported by Murakami *et al.* (1998) and Razis *et al.* (2017). Brilhante *et al.* (2017) described two very important *Moringa* bioactive glycosides (Niazimicin and Niazinin) in their comprehensive review based on 113 references. Earlier Guevara *et al.* (1999) presented evidence on the function of niazimicin present in *M. oleifera* from Philippines as an inhibitor against the two-stage mouse tumorigenesis. It decreases the proliferation and invasion of cancer cells (Jung, 2014). Crude aq. extract of Moringa (Tiloke *et al.*, 2013) and ethanolic and methanolic extracts of Moringa foliage are reported to be antiproliferative and inhibitory to cancer cells (Lea *et al.*, 2012).

### 13) Anti-Colon-Cancer

Shousha *et al.* (2019) reported that silver nano-particles (AgNPs) into *M. oleifera* extract enhanced the *in vitro* antioxidant efficiency. Polyphenolic compounds and their scavenging against free radicals are effective against the growth of colon cancer cells. A literature survey on the subject is presented by Christiano and Smarandache (2019) and they rated it as possible anti-cancer treatment with further future research.

**14) Breast and colo-rectal cancers**

Al-Asmari *et al.* (2015) reported anti-malignant properties of *M. oleifera* leaves collected from the Saudi Arabian region. The survival of breast and colorectal cancer cell lines was significantly reduced by extracts of leaves and bark. Previously, Eugenol (present in *M. oleifera* bark) was shown to be active against breast cancer (Al-Sharif *et al.*, 2013). The role of Eugenol in cell apoptosis was reported by Lacroix *et al.* (2006). According to Charoensin (2014) *M. oleifera* leaves possess antioxidant activity as well as cytotoxic and chemopreventive properties, *in vitro*, in three types of cancer cell lines: hepatocarcinoma, colorectal adenocarcinoma and breast adenocarcinoma.

**15) Hypocholesterolaemic activity**

Ghasi *et al.* (2000) reported hypocholesterolaemic effects of crude leaf extract of *M. oleifera* in Wistar rats fed with high-fat diet. B-sitosterolin in seed oil is considered to have a role in cholesterol metabolism by lowering the low-density lipoprotein (LDL) cholesterol in blood (Baeza-Liménez *et al.* (2022)

**16) Hemoglobin promoting activity**

Safitri and Retnaningsih (2021) reported significant increase in hemoglobin in pregnant rats due to *Moringa* leaves probably due to high iron content in leaf.

**17) Anti-urolithiasis effect**

Karadi *et al.* (2008) have shown anti-urolithiatic effects of *M. oleifera* root bark.

**18) Testicular damage and fertility of females**

Nayak *et al.* (2020) reported that administration of *M. oleifera* ethanolic extract of leaves mitigated cyclophosphamide-induced testicular toxicity by improving blood and intra-testicular hormonal milieu as well as modulating the expression of genes pertaining to Sertoli and spermatogonial cells in adult Swiss albino mice. Previously, Nayak *et al.* (2016) had earlier reported that combining *M. oleifera* extract with cyclophosphamide increased the sperm density, motility and reduced head defect and DNA damage. Aq. extract of *Moringa* is also reported to protect against Oxidative DNA damage (Singh *et al.*, 2009). According to Albasher *et al.* (2021) *M. oleifera* extract ameliorated testicular damage induced by lead acetate. It is a good treatment to lead toxicity. *Moringa* may alleviate erectile dysfunction (ED) in mice due to polyphenols which improve nitric oxide production and testosterone level. Dr. Shahzad Basra (video on youtube) reiterated that it may overcome ED due to its contents of l-citrulline, l-arginine, and Co-enzyme Q-10, Zn and Se. Its aqueous extract (Zade *et al.*, 2013) in male albino rats and hexanoic extract in male mice (*Mus musculus*) (Cajuday and Pocsidio, 2010) are reported to increase sexual desire and sperm count. It may improve fertility in female rats.

**19) Anti-fertility activity**

The root extract promotes inhibition of the uterus for implantation of the fertilised eggs (Prakash *et al.* (1987). The aqueous and 90% ethanol leaf extracts affected the foetal development and completely aborted implantation (Shukla *et al.*, 1988; Nath *et al.*, 1992). A high dose of the root extract (600 mg/kg) has anti-progestational activity in rats (Shukla *et al.*, 1988). It also reduced the protein concentration for the formation of the uterus (Prakash *et al.*, 1988).

**20) Antihypertensive**

Juice of *Moringa* influences blood pressure (Dahot, 1988). The leaves contain mustard oil glycosides, thiocarbamate glycosides and nitriles that may help in bringing down blood pressure (Faizi *et al.*, 1995).

**21) Cosmetic use**

The oil derived from *M. oleifera* seeds is widely utilized in cosmetic (Caceres and Lopez, 1991). Leaf extract is rich in phenolic compounds. Niziot-Tukaszevska *et al.* (2020) reported that up to 5% extract concentration of leaf shows positive effects on cell metabolism and may contribute to reduction of oxidative stress in cells. Its addition in cosmetics may improve their safety and reduce skin irritation.

**22) Disinfectant**

Seed extract of *M. oleifera* has potential of a disinfectant (Bichi *et al.*, 1995).

**23) Biopesticide**

Fuglie (1999) described Leaves as possible source of Biopesticide.

**24) Anti-allergic activity**

Abd Rani *et al.* (2019) reported anti-allergic activity of *Moringa* extracts by in both the early and the late phases of allergic reactions presumably due to the presence of several active compounds in various plant parts. Such compounds include - Ethyl-(E)-undec-6-enovate; 3, 5, 6 –trihydroxy-2-(2, 3, 4, 5, 6 –penta hydroxyphenyl)-4H-

Chroman-4-one; Quercetin; Kaempferol; B-Sitosterol-3-O-glucoside; Oleic acid; Glucomoringin; 2, 3, 4 – trihydroxybenzaldehyde and Stigamasterol

### 25) Cardiovascular disorders

The data collected on rats (Male Wistar Kyoto) support the use of orally administered *M. oleifera* seeds in diet against cardiovascular disorders associated with oxidative stress and hypertension (Randriam Boavonjy *et al.*, 2017). Its extracts alone or in combination with extracts from other plant like *Peristrophe bicalyculata* is reported to act against diabetes in alloxan-induced diabetic wistar rats and potential efficacy in CVDs (Iwara *et al.*, 2014).

### 26) Diarrhea

The antispasmodic and antimicrobial activities of roots have been reported for treatment of diarrhea (Cáceres *et al.*, 1992).

### 27) Anti-obesity

Methanolic extract of *M. oleifera* has been reported to be beneficial to the weight management (Bais *et al.*, 2014) of fat induced obese rats. Metwally *et al.* (2017) have concluded that it is reasonable to assume that antiobesity, Antiatherogenic and antidiabetic properties of *M. oleifera* are achievable and this plant could be a good therapeutic candidate in this respect. In high fat-induced rats significant amelioration of obesity was considered as a result of their antioxidant potential. Nahar *et al.* (2016) have reported that consumption of *M. oleifera* may modulate obesity in rats when administered with dose of 50 mg/ day/ rat orally for 35 days with high fat diet. The anti-obesity activity of *M. oleifera* was considered by Redha *et al.* (2021) to be owing to the improvement of lipid profile, body mass and significant regulation of genes associated with adipogenesis, glucose uptake, insulin resistance and hormones such as leptin, vaspin, resistin and insulin.

### 28) Diuretic activity

Administration of seed infusion has been reported to show diuretic activity and inhibition of carrageenan-induced edema (Cáceres *et al.*, 1992). In swiss albino rats, alcoholic extract of *M. oleifera* leaves produced dose dependent diuretic action with doses of 50, 100 and 200 mg/kg body weight (Tahkur *et al.* 2016).

### 29) Immune Disorders

*M. oleifera* is active in immune disorders (Xiao *et al.*, 2020). It is found to stimulate immune system (Sudha *et al.*, 2010).

### 30) Antinociceptive and anti-inflammatory effects

Ethanol extracts of *M. oleifera* leaves can be a potential source of anti-inflammatory agents compared to acetone extract and the standard drug (Padmalochana, 2018). Sulaiman *et al.* (2008) evaluated the antinociceptive and anti-inflammatory effects of the aqueous extract of the leaves of *M. oleifera* in laboratory animals. The extract exhibited significant antinociceptive activity and they confirmed the traditional uses of *M. oleifera* in the treatment of pain and inflammation. Cáceres and Lopez (1991), Cáceres *et al.* (1992), Ezeamuzzle *et al.* (1996), Guevara *et al.* (1996), Mehta and Agrawal, 2008, Ndiaye *et al.* (2002), Mahajan and Mehta (2008), etc. are some of the important publications on the subject. Alkaloids of Moringa have activity similar to that of ephedrine and may be useful in therapy for asthma. Moringine shows bronchiole relaxation activity (Kirtikar and Basu, 1975).

### 31) Analgesic Activity and Local Anaesthetic Activity

Alcoholic extract of seeds of *M. oleifera* exhibited potent analgesic activity (Sutar *et al.*, 2008). The root bark has also exhibited significant local anaesthetic activity (Bandana *et al.*, 2003).

### 32) Anti-ulcerogenic

Protection against gastric ulceration was significantly evident in rats which were treated with indomethacin to produce ulceration. Protection was in dose dependent manner of the leaf extract (100-400 mg/ kg BW) possibly due to tannins and flavonoids (Dahiru *et al.*, 2006). Cáceres *et al.* (1991) have asserted that the presence of flavonoids in *Moringa oleifera* should offer some protection in ulcer development.

### 33) Anti-asthmatic effects

Agarwal and Mehta (2008) reported that alkaloids present in *M. oleifera* act like ephedrine and may treat asthma. These alkaloids unwind bronchioles. Bronchial asthma has successfully been treated utilizing seed kernel of Moringa and furthermore enhanced respiratory capacities (Agarwal and Mehta, 2008).

### 34) Hepatoprotective

Hepatoprotective effects of hydroethanolic extract of *Moringa* foliage are reported by Fakurazi *et al.* (2012) and Ujah *et al.* (2013). Pari and Kumar (2002) have reported hepatoprotective activity of *M. oleifera* ethanolic extract in anti-tubercular drug-induced (isoniazid, rifampicin and pyrazinamide) liver damage in rats. Medicinal properties are attributed to the bioactives present in *Moringa*. However, the underlying mechanisms remain unclear (Ma *et al.*, 2020).

### 35) Gum exudates of *M. oleifera*

The gum released from stem of *M. oleifera* is initially white in colour which on long exposure turns to reddish brown or brownish black in colour (Sonica *et al.*, 2020). Gum exudates of the *Moringa* tree are of immense medicinal importance in treatment of some diseases like asthma, dysentery and intestinal cancer (Gupta *et al.*, 2018). It is useful as stabilizer, binder, mucoadhesive (Sonica *et al.*, 2020). It is nutritive – contains vitamins B<sub>2</sub>, B<sub>6</sub> and C, Mg, Ca, Fe, proteins, carotenoids and provitamin A (Mehta *et al.*, 2011b).

### 36) Seed oil quality

Seed oil of *M. oleifera* is light yellow in colour and has nutty flavour. It is up to 40.39% of the seed in Sindh accession of the plant (Anwar and Rashid, 2007). According to Anwar *et al.* (2007), the composition of sterol in *Moringa* oil comprises  $\beta$ -sitosterol (major component) and also stigmasterol, campesterol and D5-avenasterol. Thermal stability of *Moringa* oil is higher than soybean, sunflower, canola, cotton seed oils. It can be used in the formation of vanaspati, margarine, bakery shortening, as salad oil, cooking oil, for frying of potato chips and as frying fat in industry and restaurants (Anwar *et al.*, 2007; Nadeem and Imran, 2016). It is high in oleic acid content and may be new source of edible oil. According to Barkat and Ghazal (2016) seed oil contains predominantly oleic acid (Table 8) – similar to olive oil. A seed ointment had an effect similar to neomycin against *Staphylococcus aureus* pyoderma in mice. Seed oil of a sister species, *Moringa concanensis*, collected from Tharpakar, also has oleic acid in predominance (68%) and palmitic acid 11.4% (Manzoor *et al.*, 2007).

### 37) Seed cake

Proteins may be extracted from the seed cake for water purification. The addition of pressed cake in diet of sheep with soybean has been shown to boost ruminal fermentation and weight gain in animals directly proportional to the supplied dose (Ben Salama and Makkar, 2009).

### 38) Seeds as fertilizer

Seeds may be used as fertilizer (Martin *et al.*, 2013). The seed husk may be used in ethanol production.

### 39) Bio-preservation

Leaf extract of *M. oleifera* has some potential in bio-preservation of giant freshwater prawn, *Macrobrachium rosenbergii* (Karim *et al.*, 2018). Ayirezang *et al.* (2020), have opined that *Moringa* extracts may be used as bio-preservation agents for prolonging the shelf life of food products.

Table 8. Fatty acid composition (%) of seed edible oil (Source: Barkat and Ghazal, 2016) from Egypt. The trees were grown in three different areas.

Fatty acid	Localities of Egypt			
	M. Asuit	M. Oraby	M. Monofya	Olive oil*
Palmitic acid	6.09	5.66	6.44	7.5-20
Palmitoleic acid	1.80	1.43	1.92	3.0-3.5
Margaric acid	0.08	0.09	0.09	0- 0.3
Heptadecanoic acid	0.10	0.06	0.06	0- 0.30
Stearic acid	7.94	4.79	7.12	0.5-5.0
Oleic acid*	73.30	79.58	73.51	55.0-83.0
Linoleic acid	0.59	0.58	0.59	3.5-21.0
Linolenic acid	1.7	0.15	0.17	0.0-1.0
Arachidic acid	5.1	1.57	4.71	0.0- 0.6
Gadoleic acid	1.18	3.16	2.74	0.0- 0.40
Behenic acid	3.62	2.89	2.62	0.0- 0.20

\*, Egyptian standard (2005 for olive oil).

### 40) Biodiesel

Biodiesel production from less familiar and unconventional oils including *Jatropha*, *Moringa*, *Pongamia*, and tobacco have received greater attention in recent years (Rashid *et al.* 2008; Kivevele and Huan 2015). A survey conducted on 75 Indian plant species have concluded that fatty acid methyl esters (FAMES) of *Moringa* seed oil

meet all the main specifications of biodiesel standard of Germany, Europe, and the United States (US) (Azam *et al.* 2005). *M. oleifera* seeds contain 33–41 % (w/w) oil, known as “ben oil”. It possesses significant resistance to oxidative degradation (Rashid *et al.* 2008). Moringa seeds oil is a potential candidate for biodiesel production (Rashid *et al.*, 2008). Azad *et al.* (2015) reiterated the prospect of *M. oleifera* seed oil as a sustainable biodiesel.

#### 41) Developing effervescent granules of *Moringa*

Effervescent granules were developed from leaves of *M. oleifera* by Rani *et al.* (2021) with the following formula which was evaluated best through organoleptic and sensory examination – acceptable in terms of colour, taste, aroma and texture.

*Effervescent granule formula:*

*Moring dry leaves powder 2g + Citric acid 2.3g + NaHCO<sub>3</sub> 2.70g + Xanthan gum 0.5g + sucrose 10 g + Stevia 1.6 g + Sodium benzoate 0.05 g + Poloxamer188 0.125g + Meltodextrin 2.5 g + Melon flavour 2.075 g + Strawberry flavour 1g + Pure water 20 mL.*

The Plant was harvested from Bogo village, Boganegoro, East java. The leaves were sorted, washed, drained and dried by aerating in the shade at 25-35°C and humidity 50% until moisture was 10%. Wet granulation method was employed with above formula and granules were sieved through mesh No. 10 sieves and dried in oven at 50% till moisture was 3-5%.

#### 42) *Moringa* and hair loss

In vitro application of *Moringa* at the dose of 25 to 100 µg/ mL, provided significant protection from Gentamicin-induced hair cell loss. With dosage above 100 µg/ mL it provided near complete protection. Assay of *Moringa* extract by Broderick *et al.* (2021) demonstrated suppression of ROS, preservation of cytochrome oxidase activity, reduction in Caspase production and prevention of cell apoptosis *Moringa* extract protected against aminoglycoside -induced hair cell death also (Broderick *et al.*, 2021).

#### 43) The psychological and spiritual properties of *M. oleifera*

Traditional therapeutic, psychological and spiritual effects of *M. oleifera* according to the principles of traditional Ayurvedic and Chinese medicine systems are discussed by Meireles *et al.* (2020). According to them, *M. oleifera* penetrates the deep layers of body tissues and particularly into the bone marrow. It purifies blood, removing impurities, toxins, parasites and metabolic wastes. *Moringa oleifera* strongly influence personality. Kaur *et al.* (2015) suggested it to have anti-depressive and anxiolytic effects. Drue and Minor 2018) considered it to be an adaptogenic and anti-stress plant. According to ancient ayurvedic physicians, it contributes to the feeling of certainty, courage and fearlessness. Its root is known to enhance feelings of serenity and balance in adverse uncertain situations.

The seeds of the plant were used as a remedy for depression in ancient India. The seeds were reported tonifying and were believed to renew the spirit and reinvigorate the body as well as the mind and emotions (Warrier *et al.*, 2010). The flowers were believed to be beneficial in traumatic memories. They are said to encourage positive thinking (Warrier *et al.*, 2010).

#### 44) *Moringa* in climate-change-mitigation and global warming

*M. oleifera* can play a significant role in soil and water conservation and in mitigating climate change. It is well adapted to weather, soil and other environmental adversities (Daba, 2016). The growth of the plant even in dry conditions is considered to act as good sink for CO<sub>2</sub> absorption and utilization. According to a study, the rate of CO<sub>2</sub> absorption by *Moringa* is 50 times higher than that of general vegetation (Villfuerte and Villfuerte-Abonal, 2009).

Another important study on *Moringa* and global warming is from Muriel (2010). According to him, one person emits 320 kg of CO<sub>2</sub> per year and it takes 23 Japanese cedar trees 50 years to absorb this amount of CO<sub>2</sub> but it takes two *Moringa* trees only two years to absorb this amount. Furthermore, If a family emits 2300 kg of CO<sub>2</sub> per year, it takes 160 Japanese cedar trees 50 years to absorb this amount of CO<sub>2</sub>. On the other hand, it takes 10 *Moringa* trees two years to absorb this amount of CO<sub>2</sub>. *Moringa* is thus a useful tool to sequester more CO<sub>2</sub> and fight global warming. Planting *Moringa* will, hopefully, mitigate the impacts of climate change (Muriel, 2010).

#### 45) Water purification

Seed powder with or without husk has coagulant, flocculant, water softening and disinfectant effects in surface water, shallow water and groundwater purification (Santos Bazanella *et al.*, 2008; Sanchez- Martin *et al.*, 2010; Pritchard *et al.*, 2009, 2010; Poumaye *et al.*, 2012; Teh and Wu, 2014; Velázquez – Zavala *et al.*, 2016; Muyibim and Evison, 1994; Pollard *et al.*, 1995; Nda Bigengesser and Narasiah, 1998; Kalogo *et al.*, 2000; Anwar *et al.*, 2007, Sanchez-Martin *et al.*, 2010). Good coagulation activity of seeds on the basis of microbial and physico-chemical

studies was also recently reported by Osarughe *et al.* (2020) in case of industrial and Kitchen effluents. Adding seed powder may remove 90-99% of the bacteria present in the water (Omotesho *et al.*, 2013). After adding powder to water, it is stirred for 10-15 minutes and left for an hour or until water becomes clear and impurities sink in the bottom. Water should be filtered and boiled before drinking (Sutherland *et al.*, 1989; Jahn *et al.*, 1986; Mahmood *et al.*, 2010). Higher is the turbidity of water and temperatures, higher is the coagulant efficacy in an alkaline medium (Sanchez- Martin *et al.*, 2010; Pritchard *et al.*, 2010). Two-step purification of *M. oleifera* had significantly better turbidity removal as compared to singly-step purification (Sanchez- Martin *et al.*, 2010). It is reported to remove Calcium, Magnesium, Iron, Strontium, Aluminium (Bichi, 2013), Cadmium (Abedini and Alpour, 2015), nitrates (Rezende *et al.*, 2016), *textile dyes* (Beltran-Heredia *et al.*, 2012b) and detergents (Beltran-Heredia *et al.*, 2012 a).

Bark has been reported to remove Nickel, lead, Sodium, Potassium, Calcium, and Magnesium (Reddy *et al.*, 2010 b) and leaf has been used to remove lead (Reddy *et al.*, 2010 a). Moringa mixed with activated carbon has been used to remove Copper, Nickel, Zinc, Cadmium, Chromium, Lead, etc. (Kalavathy and Mirinda, 2010; Bhargave *et al.*, 2015).

Kalibbala *et al.* (2009) reported that *M. oleifera* can remove trihalomethane (THM) precursors and iron from drinking water successfully. Fluoride from water is removed by interaction between *M. oleifera* and negative charge of the fluoride ion through ion adsorption mechanism. Fig. 3 describes interaction between negatively charged colloids or stable particles with cationic proteins of *M. oleifera* and resulting floc formation and sedimentation of colloids (see Saini *et al.*, 2016). Gassenschmidt *et al.* (1995) isolated a flocculating protein from *M. oleifera*. Moringa also provide protection against Arsenic. The review published by Idris *et al.* (2016) based on 81 references present comprehensive data on seeds of *M. oleifera* in environmental application and describes from various references that seeds of this species may remove Arsenic, Cadmium, Argentum, Coupper, Lead, Chromium, Zinc and Mangnese in substantial amounts. They can remove pollutant like sodium lauryl sulphate from aqueous solution, humic acid and dyes, colour from distillery spent wash, water hardness, suspended solids, turbidity from palm oil mill effluent waste etc. (Idris *et al.*, 2016).

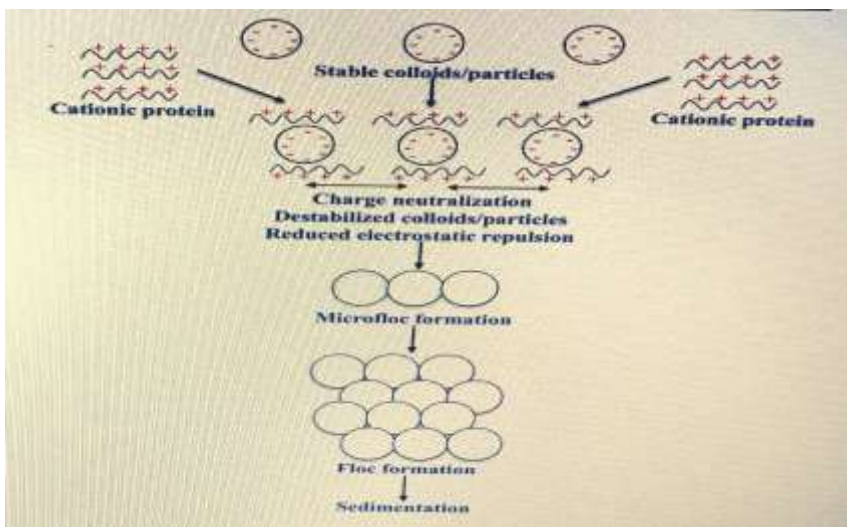


Fig. 3. Mechanism of Watercoagulation and sedimentation using *M. oleifera* cationic proteins. (Source: Saini *et al.*, 2016).

Sinha *et al.* (2016) compared ethanolic extracts of *M. oleifera* and *Azadirachta indica* for water purification potential of the two species. Ethanolic extract of leaves of these species were found by them greatly efficient in the clarification and sedimentation of total solids of tannery wastewater sample. After 5-day incubation of water with the extract sample treated with extract reduction in water quality parameters (pH, EC, Cl, SO<sub>4</sub>, hardness and TDS (Table 9). The dissolved oxygen (DO) increased by 83.33 % in *M. oleifera* and 133.33% under *A. indica* influence. EC and SO<sub>4</sub> were almost equally reduced in the two treatments but chlorine was significantly more reduced with *A. indica*. The pH of water was slightly more acidic after treatment with *M. oleifera*. Summarizing the results of Sinha *et al.* (2016), the overall effects with two species were more or less similar but DO was better elevated with *A. indica*.

Table 9. Water parameters of untreated and *Moringa oleifera* and *Azadirachta indica* treated tannery wastewater (data from Sinha *et al.* (2016) after 5days of treatment. % Promotions or reductions over untreated control were calculated by us to compare the two plant species. Formula = Treated –Untreated / Untreated \* 100.

Parameters	Untreated (control)	<i>Moringa</i> treated	% Promotion or Reduction	<i>Azadirachta indica</i> treated	% Promotion or Reduction
pH	7.30	4.85	-33.56	5.14	-29.59
DO	2.40	4.40	83.33	5.60	133.33
EC	2.34	0.897	-61.67	0.896	-61.71
Cl	65.32	62.48	-4.35	29.82	-54.34
SO <sub>4</sub>	70.0	10.40	-85.14	11.60	-83.43
Hardness	2380	2040	-14.29	1760	-26.05
TDS	382.8	192.0	-49.84	184.2	-51.88

#### 46) *Moringa* forage for feeding animals

Hanuk (2018) reviewed the use of *M. oleifera* leaves as forage for feeding animals, researched by various workers on cattle, buffalo, broilers, laying hens and rabbits by supplements to their diets. It improved growth of cattle and buffalo and rabbits, improved egg production in hens. The inclusion of seed meal at the rate of 7.5% was not found lethal to the experimental broiler chicks (Annongu *et al.*, 2014). The supplementation of *Moringa* may, play a role in the immunity building, sound health and poultry production performance (Mahfuz and Piao, 2019). No lethal dose was discovered for *Moringa* supplement in rats (Villarruel-López *et al.* (2018). No significant difference in genotoxic parameter of rats supplemented with *Moringa* was found. Nutrition value of leaves is described along with traditional medicinal use of the plant parts in detail by Agbogidi and Ilondu (2012).

High level of crude protein and the index of digestible protein in the intestine (PDI) make the *Moringa* leaf a protein supplements to animals for highly productive cattles (Makkar and Becker, 1996). In Nicaragua, good results have been obtained with *Moringa* leaves added with molasses and sugarcane straw (Radovich, 2011).

Nathaneil (2021) have assessed the effectiveness of *Moringa oleifera* and *M. stenopetala* leaves on growth, blood and gut microbiota in broiler chicken (Cobb500 broiler chicken from Kenchic Ltd. Kenya). The leaf powders from the two species were potent promoters in terms of weight gain. Similar results were obtained with ethanolic and aqueous extracts. There was no negative effect on haematological parameters. The chickens which received leaf powder of the two plant species had significantly low count of coliform. They suggested the leaf powders of *M. oleifera* and *M. stenopetala* as alternative and safer broiler chicken feed supplements. Further validation and authentication of the use of two species is, however, needful.

#### 47) *Moringa Oleifera* active against *Anopheles gambiaes* and *Aedes aegyptii*

*M. oleifera* also inhibits growth of larvae of anopheles gambiaes (Chuang *et al.*, 2007; Prabhu *et al.*, 2011). *Moringa* activity against *Aedes aegyptii* (vector of dengue virus) owing to its contents of  $\beta$  - amyryn,  $\beta$ -Sitosterol, Kaempferol and Quercitin (Pontual *et al.* (2012).

#### 48) *Moringa* in poverty alleviation

Ometesho *et al.* (2013) indicated potential of *M. oleifera* tree for poverty alleviation and rural development. It can be used to overcome malnutrition (Halder and Kosankar, 2017).

#### 49) *Heliobacter pylori*

*Moringa oleifera* Lam. is rich in fibers and isothiocyanates which have anti-bacterial activity. It may help to rid off *H. pylori*, the bacteria causing gastritis and gastric cancer (Paikra *et al.*, 2017).

#### 50) Antiepileptic activity

Amrutia *et al.* (2011) reported that methanolic extract of *M. oleifera* leaves to exhibit potent anticonvulsant activity at the dose level of 200mg / kg and 400 mg/kg administered intraperitoneally when diazepam and phenytoin were used as reference standard.

#### 51) Oral hygiene

*M. oleifera* is used for oral hygiene (Jose *et al.*, 2011). It is used to mineralize enamel and dentin in patients having erosive and wasting disease of teeth (Khalaf *et al.*, 2016). It is antimicrobial and works against dental plaque (Yadav *et al.*, 2021).

#### 52) Wound-closure and tissue regeneration

Improvement in tissue regeneration, decrease in wound size, and remarkable anti-proliferative and anti-migratory effects on normal human dermal fibroblasts have been reported (Cácares *et al.*, 1992; Lambole and Kumar, 2012). The antibacterial efficacy of aqueous and ethanol extracts of fresh and dried leaves of *Moringa oleifera* against



several pathogens isolated from wound and faeces, indicated potential their in the treatment of wound infection and typhoid fever (Dike-Nduddin *et al.*, 2016). Antipyretic and wound healing properties of ethanolic and ethyl acetate leaf extracts of *Moringa oleifera* have also been reported by Hukkeri *et al.* (2006).

### 53) Lead toxicity and Moringa

Lead is dangerous pollutant and shows nephrotoxic effects in humans. Curing effects of leaf extract of Moringa have been recorded on lead toxicity (Mohamed *et al.*, 2020) in rats. Further studies are, however, suggested to determine safety for prolonged use of its leaves.

### 54) Osteoporosis

The *Moringa* flavonoids increase bone density, which allows to prevent osteoporosis (Nijveldt *et al.*, 2001).

### 55) Urinary tract infections

Shaw and Jana (1982) reported efficacy of *M. oleifera* in lower urinary tract infections.

### 56) Toxicity due to Moringa

Steroids, alkaloids, tannins and saponions are considered poisonous compounds of Moringa. It is toxic at certain doses and overuse can cause genotoxicity (Liu *et al.*, 2021). Intake of root bark juice may cause acute skin inflammation and dermatitis (Alagesa Boopathi (2019). Hydroethanolic extract from Moringa cultivated in Selangor, Malaysia, was lethal to female ICR-mice if given in doses higher than 2000mg/kg. At this level the extract was reported to cause mild anaemia, stress leucogram and mild to moderate hepatonephrotoxicity (Aliyu *et al.*, 2021). They considered regular and repeated dose at 1000mg/kg to be unsafe. However, lower doses and /or administration for short time could be used safely for medicinal purpose. Adepapo *et al.* (2009) have reported that plant is relatively safe for nutritional and medicinal uses. Horas *et al.* (2021) have reviewed and characterized *M. oleifera* as a functional food and natural food additive. They, however, opined that many reports are inconclusive, and some data are even contradictory. It was probably the reason for its commercialization prohibition in Brazil in 2019 (Horas *et al.*, 2021). Earlier, Farooq *et al.* (2012) also suggested for its legitimate appraisal in modern medicine. Moreover, the content of Arsenic in San Pedro (Mexico) sample was reported to be higher than the permissible limit (Valdez-Solana *et al.* (2015).

### 57) Future risk to *Moringa oleifera*

The utility of *M. oleifera* is immense which may lead to its over-exploitation. Therefore, Gupta *et al.* (2018) have suggested the conservation of the tree and its afforestation on large scale. In planted populations of *M. oleifera* in Sanja, North Gondar, Ethiopia by farmers, Gedefaw (2015) have reported Moringa aboveground biomass varying with size between 9.788 and 197.485 ton per Ha and belowground biomass between 1.96 and 39.49 ton / ha. He also reported on carbon stocks in the aboveground biomass varying from 4.894 to 98.742 ton per ha in the plots studied.

Of course, there is need of raising awareness about the merits of Moringa to the people who should undertake plantation of this tree around houses and in the field. The farmers should come forward planting this plant which is fast-growing and beneficial in so many ways.

As discussed above in the paper, *M. oleifera* is an exceptionally important multipurpose tree. Its usefulness in therapeutics and other areas cannot be overemphasized but still regressive research is needed on various aspects. More comprehensive studies related to the phytochemicals should hopefully enhance pharmaceutical exploration in the field of cancer prevention (Al-temimi *et al.*, 2017). Moringa extracts provide unlimited opportunities for discoveries of new drugs owing to a variety of chemicals present in them (Sridharan *et al.*, 2011). Data from real clinical trials are, however, meagre and studies on extracts activity have generally been made *in vitro* (Lui *et al.*, 2021). Since there is still some lack of data on the pharmacology, toxicity, agricultural economy and dietary benefits of its constituents, its extracts require further evaluation. There are, however, many studies confirming to its usefulness in many ailments in human populations owing to its richness in antioxidants with rational roles as prophylactics and therapeutic actions. Although it has multifaceted pharmaceutical potential and some breakthrough has been recorded in isolation, identification and structural elucidation of therapeutically bioactive principles, research is still needed to ascertain its safety in humans. Many products of Moringa in market need to be assessed for their quality and safety ((Papoola *et al.*, 2020).

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